

PCR Max Ltd™ qPCR test

Human Rotavirus B

Non structural protein 5 (NSP5)

150 tests

For general laboratory and research use only



Introduction to Human Rotavirus B

Rotavirus is a genus of double-stranded RNA based viruses of the Reoviridae family. Species within this genus have been named A to G, all cause severe diarrhoea although species A-C are the most common in humans. The linear RNA genome of this species contains 11 segments between 649 and 3538 nucleotides in length. The three layer capsid containing this genome has icosahedral symmetry and is around 80nm in diameter and has channels which extend inward to the core.

The main route of transmission is the faecal-oral route due to contamination of food and water. The virus targets the enterocytes of the intestinal villi, enters the host cell by receptor-mediated endocytosis and replicates in the cytoplasm. After replication and construction of the new viral progeny the cell is lysed resulting in viral release as well as release of intestinal fluid which results in diarrhoea containing viral particles.

After an incubation period of around 2 days, symptoms of Rotavirus infection present with vomiting, fever and diarrhoea that can last over a week. After infection antibodies to the virus can be found in an individual, subsequent infections can occur throughout life but are mainly asymptomatic. Infection is therefore most severe in young children although newborns are thought to have acquired immunity from trans-placental transfer of antibodies. Treatment is usually directed at alleviation of symptoms and mainly involves rehydration.

Specificity

The PCR Max™ qPCR Kit for Human Rotavirus B (Rotavirus B) genomes is designed for the in vitro quantification of Rotavirus B genomes. The kit is designed to have the broadest detection profile possible whilst remaining specific to the Rotavirus B genome. The primers and probe sequences in this kit have 100% homology with a broad range of Rotavirus B sequences based on a comprehensive bioinformatics analysis.

If you require further information, or have a specific question about the detection profile of this kit then please send an e.mail to pcrmaxhelp@pcrmax.com and our bioinformatics team will answer your question.

Kit Contents

- **Rotavirus B specific primer/probe mix (150 reactions BROWN)**
FAM labelled
- **Rotavirus B positive control template (for Standard curve RED)**
- **Internal extraction control primer/probe mix (150 reactions BROWN)**
VIC labelled as standard
- **Internal extraction control RNA (150 reactions BLUE)**
- **Endogenous control primer/probe mix (150 reactions BROWN)**
FAM labelled
- **Rotavirus B/Internal extraction control/endogenous control RT primer mix (150 reactions GREEN)**
Required for two step protocol only
- **RNAse/DNAse free water (WHITE)**
for resuspension of primer/probe mixes and internal extraction control RNA
- **Template preparation buffer (YELLOW)**
for resuspension of positive control template and standard curve preparation

Reagents and equipment to be supplied by the user

Real-Time PCR Instrument

RNA extraction kit

This kit is designed to work well with all processes that yield high quality RNA with minimal PCR inhibitors.

Lyophilised OneStep 2x qRT-PCR MasterMix

Contains complete one step qRT-PCR MasterMix

Pipettors and Tips

Vortex and centrifuge

1.5 ml tubes

Kit storage and stability

This kit is stable at room temperature but should be stored at -20°C on arrival. PCR Max does not recommend using the kit after the expiry date stated on the pack. Once the lyophilized components have been re-suspended, unnecessary repeated freeze/thawing should be avoided. The kit is stable for six months from the date of resuspension under these circumstances.

If a standard curve dilution series is prepared this can be stored frozen for an extended period. If you see any degradation in this serial dilution a fresh standard curve can be prepared from the positive control.

Suitable sample material

All kinds of sample material suited for PCR amplification can be used. Please ensure the samples are suitable in terms of purity, concentration, and RNA/DNA integrity (An internal PCR control is supplied to test for non specific PCR inhibitors). Always run at least one negative control with the samples. To prepare a negative-control, replace the template RNA sample with RNase/DNase free water.

Dynamic range of test

Under optimal PCR conditions PCR Max Rotavirus B detection kits have very high priming efficiencies of >95% and can detect less than 100 copies of target template.

Notices and disclaimers

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Principles of the test

Real-time PCR

A Rotavirus B specific primer and probe mix is provided and this can be detected through the FAM channel.

The primer and probe mix provided exploits the so-called TaqMan® principle. During PCR amplification, forward and reverse primers hybridize to the Rotavirus B cDNA. A fluorogenic probe is included in the same reaction mixture which consists of a DNA probe labeled with a 5'-dye and a 3'-quencher. During PCR amplification, the probe is cleaved and the reporter dye and quencher are separated. The resulting increase in fluorescence can be detected on a range of real-time PCR platforms.

Positive control

For copy number determination and as a positive control for the PCR set up, the kit contains a positive control template.

This can be used to generate a standard curve of Rotavirus B copy number / CT value. Alternatively the positive control can be used at a single dilution where full quantitative analysis of the samples is not required. Each time the kit is used, at least one positive control reaction must be included in the run. A positive result indicates that the primers and probes for detecting the target Rotavirus B gene worked properly in that particular experimental scenario. If a negative result is obtained the test results are invalid and must be repeated. Care should be taken to ensure that the positive control does not contaminate any other kit component which would lead to false-positive results. This can be achieved by handling this component in a Post PCR environment. Care should also be taken to avoid cross-contamination of other samples when adding the positive control to the run. This can be avoided by sealing all other samples and negative controls before pipetting the positive control into the positive control well.

Negative control

To validate any positive findings a negative control reaction should be included every time the kit is used. For this reaction the RNase/DNase free water should be used instead of template. A negative result indicates that the reagents have not become contaminated while setting up the run. If a positive result is obtained the results should be ignored and the test samples repeated. Possible sources of contamination should first be explored and removed.

Internal RNA extraction control

When performing RNA extraction, it is often advantageous to have an exogenous source of RNA template that is spiked into the lysis buffer. This control RNA is then co-purified with the sample RNA and can be detected as a positive control for the extraction process. Successful co-purification and real-time PCR for the control RNA also indicates that PCR inhibitors are not present at a high concentration.

A separate RT primer mix and a real-time PCR primer/probe mix are supplied with this kit to detect the exogenous RNA using real-time PCR. The PCR primers are present at PCR limiting concentrations which allows multiplexing with the target sequence primers. Amplification of the control cDNA does not interfere with detection of the Rotavirus B target cDNA even when present at low copy number. The Internal control is detected through the VIC channel and gives a CT value of 28+/-3 depending on the level of sample dilution.

Endogenous control

To confirm extraction of a valid biological template, a primer and probe mix is included to detect an endogenous gene. Detection of the endogenous control is through the FAM channel and it is NOT therefore possible to perform a multiplex with the Rotavirus B primers. A poor endogenous control signal may indicate that the sample did not contain sufficient biological material.

Carry-over prevention using UNG (unsuitable for onestep procedure and optional for two step procedure)

Carry over contamination between PCR reactions can be prevented by including uracil-N-glycosylase (UNG) in the reaction mix. Some commercial mastermix preparations contain UNG or alternatively it can be added as a separate component. UNG can only prevent carry over from PCR reactions that include deoxyuridine triphosphate (dUTP) in the original PCR reaction. Primerdesign recommend the application of 0.2U UNG per assay with a 15 minute incubation step at 37°C prior to amplification. The heat-labile UNG is then inactivated during the Taq polymerase activation step.

Reconstitution Protocol

To minimize the risk of contamination with foreign DNA, we recommend that all pipetting be performed in a PCR clean environment. Ideally this would be a designated PCR lab or PCR cabinet. Filter tips are recommended for all pipetting steps.

- 1. Pulse-spin each tube in a centrifuge before opening.**
This will ensure lyophilised primer and probe mix is in the base of the tube and is not spilt upon opening the tube.
- 2. Reconstitute the kit components in the RNase/DNase free water supplied, according to the table below:**
To ensure complete resuspension, vortex each tube thoroughly.

Component - resuspend in water	Volume
Pre-PCR pack	
Rotavirus B primer/probe mix (BROWN)	165 μ l
Internal extraction control primer/probe mix (BROWN)	165 μ l
RT primer mix (GREEN)	165 μ l
Endogenous control primer/probe mix (BROWN)	165 μ l
Pre-PCR heat-sealed foil	
Internal extraction control RNA (BLUE)	600 μ l

- 3. Reconstitute the positive control template in the template preparation buffer supplied, according to the table below:**
To ensure complete resuspension, vortex this tube thoroughly.

Component - resuspend in template preparation buffer	Volume
Post-PCR heat-sealed foil	
Positive Control Template (RED) *	500 μ l

* This component contains high copy number template and is a VERY significant contamination risk. It must be opened and handled in a separate laboratory environment, away from the other components.

RNA extraction

The internal extraction control RNA can be added either to the RNA lysis/extraction buffer or to the RNA sample once it has been resuspended in lysis buffer.

DO NOT add the internal extraction control RNA directly to the unprocessed biological sample as this will lead to degradation and a loss in signal.

- 1. Add 4 μ l of the Internal extraction control RNA (BLUE) to each sample in RNA lysis/extraction buffer per sample.**
- 2. Complete RNA extraction according to the manufacturers protocols.**

One Step RT-PCR detection protocol

A one step approach combining the reverse transcription and amplification in a single closed tube is the preferred method.

For optimum performance and sensitivity.

All pipetting steps and experimental plate set up should be performed on ice. After the plate is poured proceed immediately to the One Step amplification protocol. Prolonged incubation of reaction mixes at room temperature can lead to PCR artifacts that reduce the sensitivity of detection.

1. **For each RNA sample prepare a reaction mix according to the table below: Include sufficient reactions for positive and negative controls.**

Component	Volume
OneStep 2x qRT-PCR MasterMix	10 μ l
Rotavirus B primer/probe mix (BROWN)	1 μ l
Internal extraction control primer/probe mix (BROWN)	1 μ l
RNAse/DNAse free water (WHITE)	3 μ l
Final Volume	15 μl

2. **For each RNA sample prepare an endogenous control reaction according to the table below (optional):**
This control reaction will provide crucial information regarding the quality of the biological sample.

Component	Volume
OneStep 2x qRT-PCR MasterMix	10 μ l
Endogenous control primer/probe mix (BROWN)	1 μ l
RNAse/DNAse free water (WHITE)	4 μ l
Final Volume	15 μl

3. **Pipette 15 μ l of these mixes into each well according to your real-time PCR experimental plate set up.**
4. **Pipette 5 μ l of RNA template into each well, according to your experimental plate set up.**
For negative control wells use 5 μ l of RNAse/DNAse free water. The final volume in each well is 20 μ l.

5. **If a standard curve is included for quantitative analysis prepare a reaction mix according to the table below:**

Component	Volume
OneStep 2x qRT-PCR MasterMix	10 μ l
Rotavirus B primer/probe mix (BROWN)	1 μ l
RNAse/DNAse free water (WHITE)	4 μ l
Final Volume	15 μl

6. **Preparation of standard curve dilution series.**

- 1) Pipette 90 μ l of template preparation buffer into 5 tubes and label 2-6
- 2) Pipette 10 μ l of Positive Control Template (RED) into tube 2
- 3) Vortex thoroughly
- 4) Change pipette tip and pipette 10 μ l from tube 2 into tube 3
- 5) Vortex thoroughly

Repeat steps 4 and 5 to complete the dilution series

Standard Curve	Copy Number
Tube 1 Positive control (RED)	2×10^5 per μ l
Tube 2	2×10^4 per μ l
Tube 3	2×10^3 per μ l
Tube 4	2×10^2 per μ l
Tube 5	20 per μ l
Tube 6	2 per μ l

7. **Pipette 5 μ l of standard template into each well for the standard curve according to your plate set-up**
The final volume in each well is 20 μ l.

One Step Amplification Protocol

Amplification conditions using OneStep 2x RT-qPCR MasterMix.

	Step	Time	Temp
	Reverse Transcription	10 mins	42 °C
	Enzyme activation	2 mins	95 °C
50 Cycles	Denaturation	10 secs	95 °C
	DATA COLLECTION *	60 secs	60 °C

* Fluorogenic data should be collected during this step through the FAM and VIC channels

Interpretation of Results

Target	Internal control	Negative control	Positive control	Interpretation
+ive	+ive	-ive	+ive	+ive
+ive	-ive	-ive	+ive	+ive
-ive	+ive	-ive	+ive	-ive
-ive	-ive	-ive	-ive	Experiment fail
+ive	+ive	+ive	+ive	Experiment fail

Internal PCR control

The CT value obtained with the internal control will vary significantly depending on the extraction efficiency, the quantity of RNA added to the RT and PCR reaction and the individual machine settings. CT values of 28 ± 3 are within the normal range. When amplifying a Rotavirus B sample with a high genome copy number, the internal extraction control may not produce an amplification plot. This does not invalidate the test and should be interpreted as a positive experimental result.

Endogenous control

The signal obtained from the endogenous control primer and probe set will vary according to the amount of biological material present in a given sample. An early signal indicates the presence of a good yield of biological material. A late signal suggests that little biological material is present in the sample.